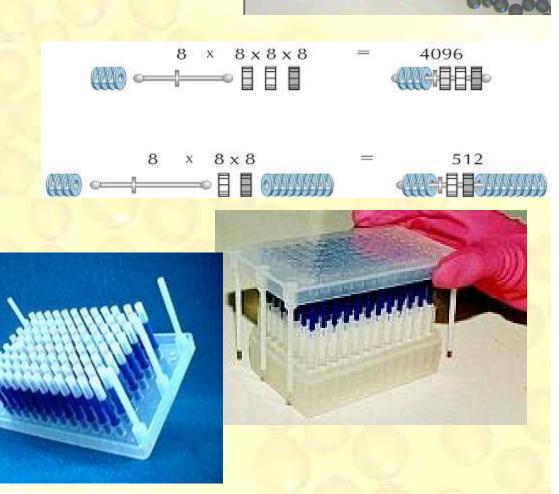
2. Support formats for SPOS

- o 2.2.4. Grafted surfaces pins
 - Any shape
 - Microtitre well format
 - Fast reaction rates
- o 2.2.5. Irregular shapes luck charms
 - Similar to beads
- o 2.2.6. Discs
 - Similar to beads for chemistry
 - Similar to luck charms for handling

- 3.1. Chemical tagging
 - o Orthogonal
 - Isotopic encoding
 - Amino acid encoding
 - o Genetic
 - Phage displays
 - o Arylhalides
 - Levels of chlorine

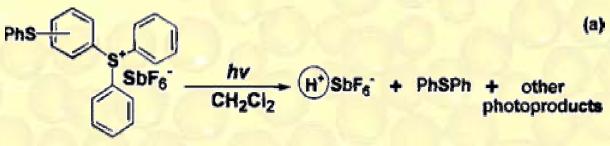
• 3.1. Physical tagging

- o Single beads / pins etc.
- o Magnetic encoding
- o Radiofrequency tags
- Colour coding

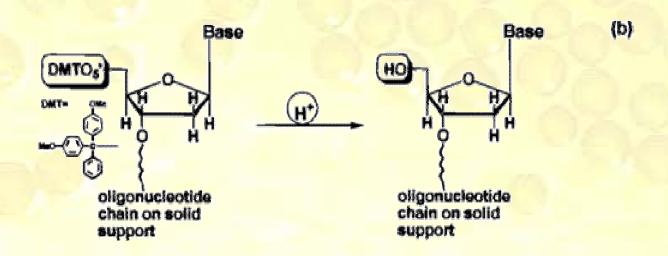


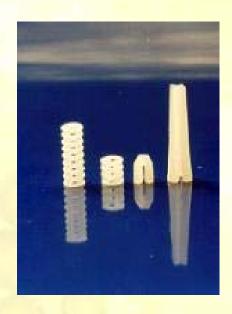
3.1. Physical tagging

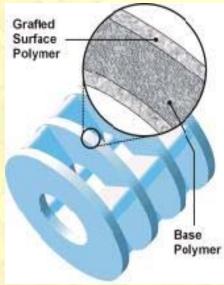
- o Unique shapes
- o Spatially addressable



SSb: triarylaulfonium PGA-P PGA

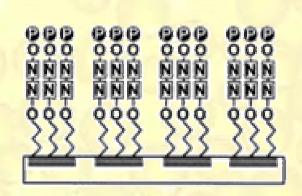


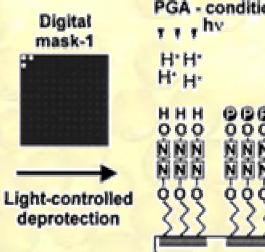


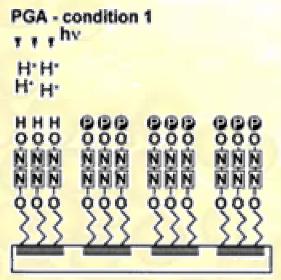


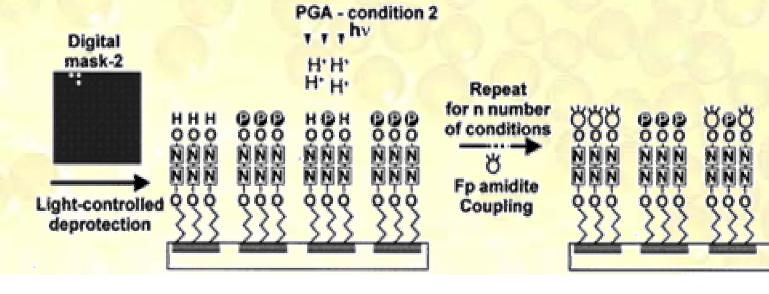
3.1. Physical tagging

o Spatially addressable





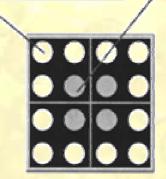






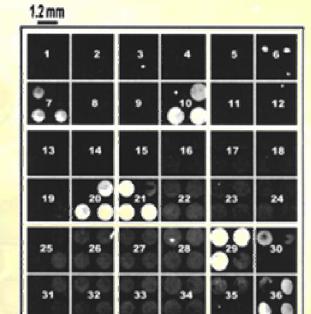


Illuminated

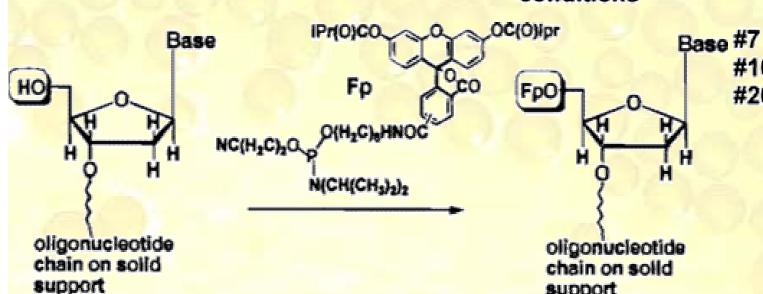


A panel of 4 PGA conditions

support

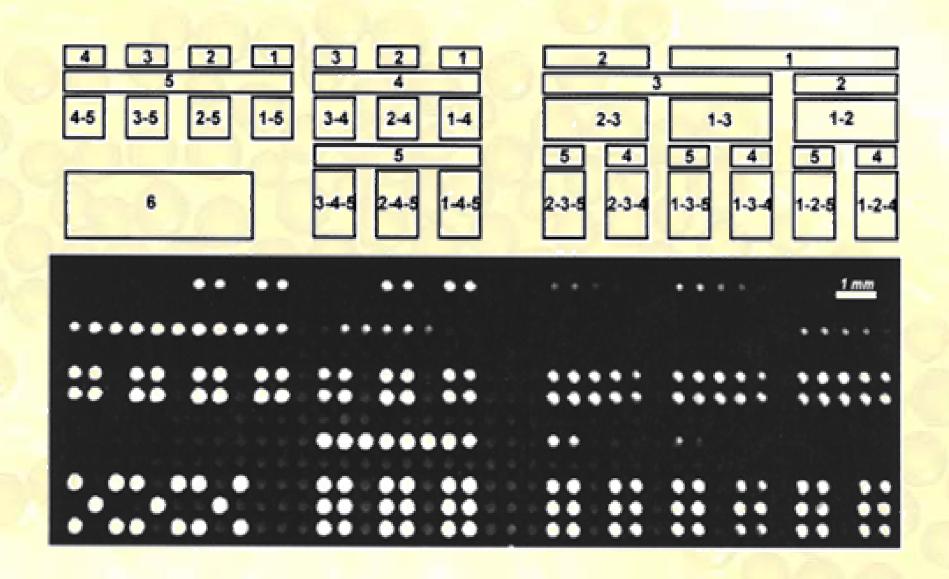


Intensity Ratio



4.2±0.2 6.6±0.5 #21 #10 5.7±0.3 6.3±0.5 #29 #36 #20 6.5±0.5 4.4±0.3

Spatially addressable



4. Separation/Handling

- 4.1. Single beads/charms
- 4.2. Pressure control
- 4.3. Split and mix
- 4.4. Tea bags
- 4.5. Magnetic separation
- 4.6. Irori tubes
 - http://www.irori.com/Technology/directed_sorting_demo. html
- 4.7. Sheets
- 4.8. Photolithographic masks
- 4.9. Cotton Strips

- 5.1. Swelling
 - o 5.1.1. Accessibility
 - Spacer length
 - o 5.1.2. Reaction kinetics
 - Gel
 - Macroporous
 - Bead Size
 - Grafting

- o Polyacrylamide
- o CPS
- o Poly(meth)acrylates
 - Gel vs Macroporous
- o Tentagel
- o Argogel

- o Tentagel
- o Argogel

Table. Swelling Volume (mL/g) for Polystyrene and Various PS-PEG Graft Copolymers at 25 C (mL/g)

solvent	PS-	TentaGel-	ArgoGel-	TentaGel-	ArgoGel-	
	NH2 a	OH b	ОН с	NH2 b	NH2 c	
			(12)			
toluene			5.2		6.2	
THF	8.0	4.5	5.8	3.9	6.4	
DCM	8.9	5.8	7.5	5.5	8.6	
DMF	8.0	4.5	6.0	4.0	5.0	
MeOH	2.8	3.8	4.7	3.0	4.9	
water	2.1	3.8	3.9	3.0	4.0	

- a Obtained from Bachem.
- b Obtained from Rapp Polymere.
- c Obtained from Argonaut Technologies.

• 5.2. Solvent compatibility

o PS

Gel vs Macroporous

Kinetics of Knorr Formation Reaction on Various Resins a resins (size)kobs (1/s)t1/2 (s)conversion (%)loading (mmol/g)b

3

NHFmoc

OMe

PS (100-200 mesh)	PS (100-200 mesh) c	PS (200-400 mesh)	PS (70-90 mesh)	ArgoGel- AM (164 μm)	TentaGel- AM (130 μm)	Champion- 1 (100-200 mesh) d	Polymer
2.1 × 10-2	7.9 × 10-3	ca. 8 × 10-2	3.9 × 10-3	ca. 7 × 10-2	ca. 6 × 10-2	ca. 4 × 10-1	k _{obs} t _{1/2} conversion loading
33	88	ca. 9	178	ca. 10	ca. 12	ca. 2	
93	71	96	96	94	91	98	
1.38	1.38	0.43	1.12	0.44	0.29	0.40	

a Experimental conditions: 0.2 M Knorr/0.4 M DIEA/0.2 M PyBOP in DCM at 25 C.

b Measured loading by Fmoc-Cl before Knorr formation (see Supporting Information for details).

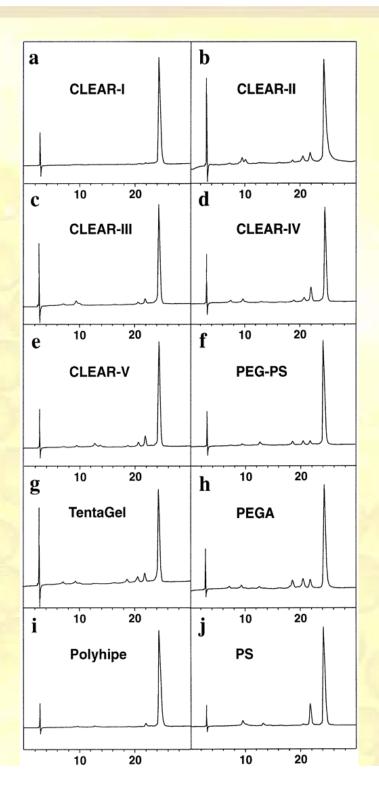
c The same experimental conditions used in previous entry but carried out in DMF instead of DCM.

d Champion-1, a type of PEG-AMPS resin that has 60% PEG content from Biosearch Technologies.8

- o Poly(meth)acrylates
 - Gel vs Macroporous

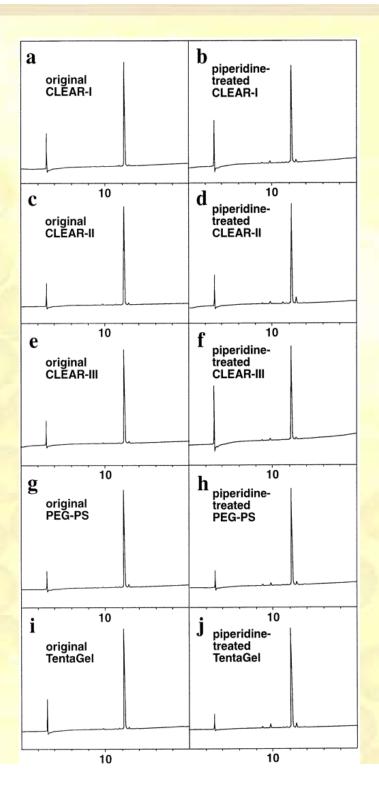
Analytical HPLC chromatogram of crude retro-ACP (74-65) amide, prepared by batchwise manual synthesis on (a) CLEAR-I; (b) CLEAR-II; (c) CLEAR-III; (d) CLEAR-IV; (e) CLEAR-V; (f) PEG-PS; (g) TentaGel; (h) PEGA; (i) Polyhipe; and (j) PS. Elutions at 1 mL/min with a linear gradient over 30 min from 17:3 to 3:1 of 0.1% aqueous TFA and 0.1% TFA in acetonitrile.

Peptides were detected at 220 nm (ordinates); time scale is in min (abscissa). The chromatogram shown in (b) was reproduced in the LC-MS mode, which indicated that the peaks preceding the major one were the des-Ile (9.4 min), des-Gln (20.5 min), and des-Asn (21.8 min) analogues of retro-ACP (74-65) amide.



o Poly(meth)acrylates

Analytical HPLC chromatogram of crude Leu-enkephalinamide, prepared by batchwise manual synthesis on (a) CLEAR-I (0.24 mmol/g); (b) CLEAR-I that had been treated with piperidine-DMF (1:4) for 9 days; (c) CLEAR-II (0.33 mmol/g); (d) CLEAR-II that had been treated with piperidine-DMF (1:4) for 9 days; (e) CLEAR-III (0.18 mmol/g); (f) CLEAR-III that had been treated with piperidine-DMF (1:4) for 9 days; (g) PEG-PS (0.18 mmol/g); (h) PEG-PS that had been treated with piperidine-DMF (1:4) for 9 days; (i) TentaGel (0.18 mmol/g); (j) TentaGel that had been treated with piperidine-DMF (1:4) for 9 days. Elutions at 0.8 mL/min with a linear gradient over 20 min from 19:1 to 2:3 of 0.1% aqueous TFA and 0.1% TFA in acetonitrile. Peptides were detected at 220 nm (ordinates); time scale is in min (abscissa).



Preparation of CLEAR Supports

SUPPORT monomer or crosslinker (mmol)	CLEAR-I	CLEAR-II	CLEAR-III	CLEAR-IV	CLEAR-V
1	9.0	3.0	8.0	12.0	13.0
2 3	9.0	3.6	4.4	50.0	50.0
4	3	7.0			
5		1700	3.0	2.0	
6	1 1		1 1	3.0	
solvent	c-hexanol	c-hexanol	c-hexanol	toluene b	toluene b
mL	17.5	16.0	20.0	12.7b	12.8b
loading a (mmol NH2/g)					
by AAA	0.26	0.30	0.30	0.17	0.13
by UV	0.29	0.25	0.27	0.23	0.20

a The loadings varied slightly from batch to batch. Incorporation of components into the final polymers did not match the initial ratios; relevant elemental analysis data in Experimental Section.

b Volume reported for these suspension polymerizations refers to organic solvent. The volume of the aqueous phase was $\sim 10 \times$ that of organic;. The amount of allylamine (2) used was high because it is soluble in both phases.

Swelling of CLEAR Supports

$$CH_{2} - (O - CH_{2} - CH_{2})_{\ell} - O - C - CH = CH_{2}$$

$$C_{2}H_{5} - C - CH_{2} - (O - CH_{2} - CH_{2})_{m} - O - C - CH = CH_{2}$$

$$CH_{2} - (O - CH_{2} - CH_{2})_{n} - O - C - CH = CH_{2}$$

$$CH_{2} - (O - CH_{2} - CH_{2})_{n} - O - C - CH = CH_{2}$$

$$CH_2 = CH - CH_2 - NH_2$$
 $CH_2 = C - C - C - CH_2 - CH_2 - N$
2 3

$$CH_{2} = \overset{CH_{3}}{\overset{\circ}{\text{C}}} \overset{\circ}{\overset{\circ}{\text{C}}} \overset{\circ}{\text{C}} + CH_{2} = CH_{2}$$

$$CH_{2} = \overset{\circ}{\text{C}} - \overset{\circ}{\text{C}} - C = CH_{2} - CH_{2} = CH_{2}$$

$$4 \qquad p \sim 9$$

bed volume (mL) of 1 g polymer

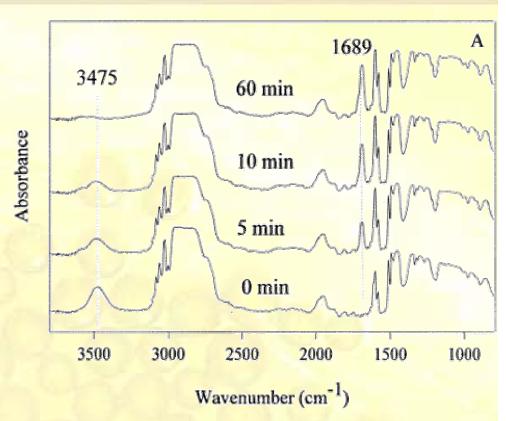
									6		
solvent	DMF	CH ₂ Cl ₂	CH ₃ CN	THF	MeOH	H ₂ O	toluene	EtOAc	t-BuOMe	hexane	TFA
CLEAR-I	8	10	6.5	6.5	7	8	5	5	3.5	3	12
CLEAR-II	5	7	4.5	4.5	5	5	4	4	4	3	7
CLEAR-III	5	7	5	5	5	5	5	4	4	3	8
CLEAR-IV	5.5	5.5	5	5	5	4	5	4.5	3	3	6.5
CLEAR-V	5	8	5	5	5	5	4	4.5	3	3	7.5

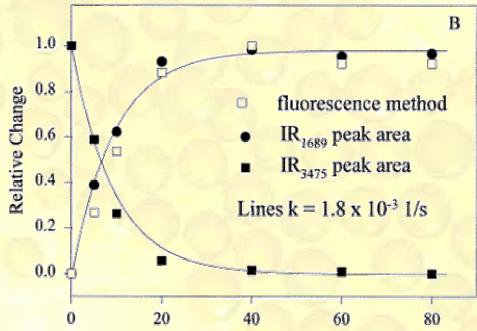
a For each set of swelling studies, the underivatized CLEAR support (0.1g) was placed in a 3-mL syringe fitted at the bottom with a porous frit. The support materials were washed 10× with the indicated solvent, and then allowed to stand for 5 min in that same solvent. Excess solvent was removed by brief suction, and the bed volume of the fully solvated support was noted. The CLEAR supports were then washed 10× with CH2Cl2 and dried [air-drying for volatile solvents; in vacuo overnight for H2O and DMF], and the procedure was repeated on the same resins with the next solvent.

o PS vs Tentagel

- Polymer represents a "solvent-like" phase
- Choice of resin depends on the nature of the reaction
- Choice of resin depends on polarity of the solvent

Reaction 1





o PS vs Tentagel

OH
$$\frac{RCOOH}{DIC/DMAP}$$

OH $\frac{RCOOH}{DIC/DMAP}$

DMF, r.t.

Product

 $\frac{3}{4}$
 $\frac{R}{4}$
 $\frac{CH_2Ph}{5}$
 $\frac{7}{6}$
 $\frac{CH_2CH_3}{6}$
 $\frac{7}{6}$

Reaction 2

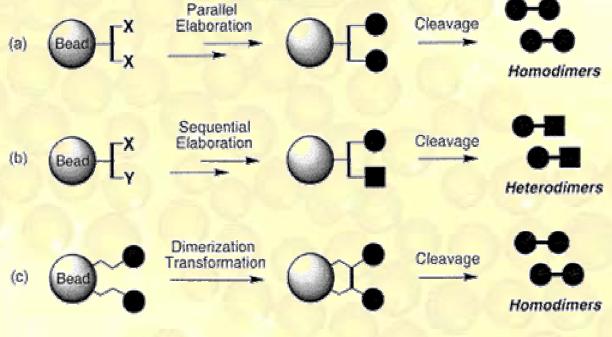
PS vs Tentagel

Reaction 4

	Comparison of Reaction Rat	es on PS- and TG-	-Based Resins rea	action scheme	
+					

Reaction	1	2 (R = 3)	2 (R = 4)	2 (R = 5)	3 (R = H)	3 (R = 3)	4 (step 2)
kPS (1/s)	4.6 × 10-4	2.2 × 10-4	4.8 × 10-4	2.0 × 10-4	3.1 × 10-3	4.1 × 10-4	1.13 × 10-4
kTG (1/s)	1.8 × 10-3	2.3 × 10-4	4.2 × 10-4	2.2 × 10-4	1.8 × 10-3	1.9 × 10-4	6.26 × 10-6
kTG/kPS	3.9	1.0	0.9	1.1	0.6	0.5	0.055

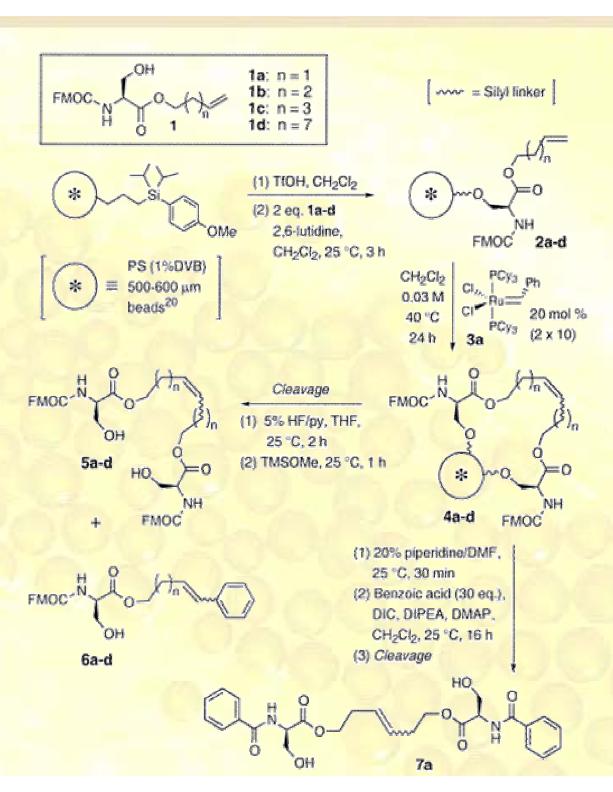
- 6.1. PS
 - o Gel
 - o Macroporous
- 6.2. Tentagel
- 6.3. Site isolation
 - o Physical meaning
 - Effect of loading levels
 - Effect of crosslinking
 - Effect of solvent
 - o Examples
 - Cyclisation
 - Dimerisations
 - Kinetics
 - Catalysis
 - Enantioselectivity



High capacity (1-2 mmol/g), lightly cross-linked (1% DVB), 500-600 m polystyrene (PS) beads.

o Examples

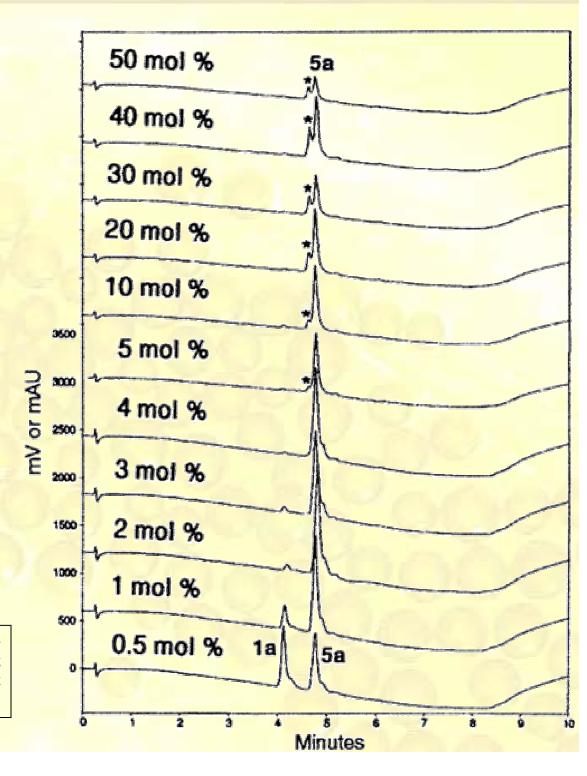
Dimerisations



o Examples

Dimerisations

Reverse-phase HPLC traces for crude intra-site crossmetathesis reactions of resin-bound monomer 1a using various catalyst loadings of benzylidene 3a. The asterisk marks the peak corresponding to styrene adduct 6a.



o Examples

- Cyclisation
- Dimerisations
- Kinetics
- Catalysis
- Enantioselectivity

7. Stability

- 7.1. Physical
- 7.2. Mechanical
- 7.3. Chemical
- 7.4. Comparison of supports

• 8.1. Acid labile

- o Wang (TFA)
- o Rink Amide (TFA)Rink Acid (very H+ labile)
- o Rink AM (95% TFA)
- o Trityl (2-chloro)

- Sulfamylbenzoyl AM (highly base stable)
- o Ketal linkers

• 8.2. Base labile

• 8.3. Photolabile

Orthogonal synthesis (Affymetrix)

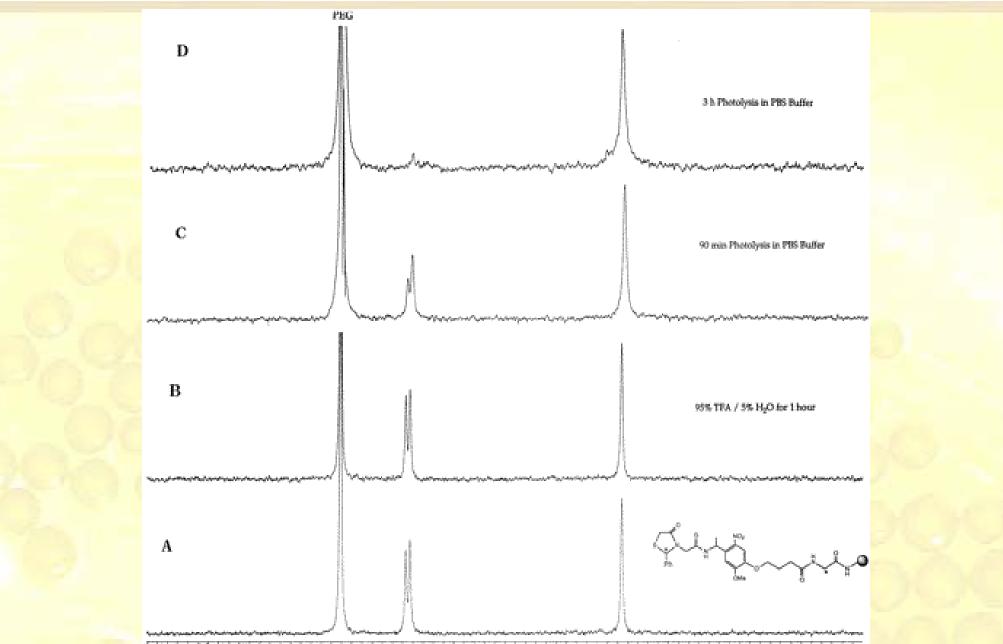
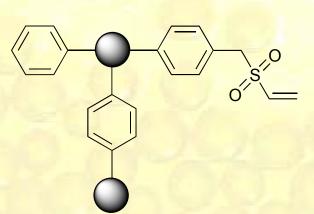


Figure 2 Gel phase ¹³C NMR trace for resin **28**. Trace A: natural abundance polyethylene glycol (PEG) is at 71 ppm, ¹³C-C₂ glycine appears at 44 ppm, and ¹³C-C₂ of the thiazolidinone appears at 65 ppm. Trace B: resin **28** after exposure to TFA/H₂O) for 1 h. Trace C: resin **28** after irradiation at 365 nm for 1 h. Trace D: resin **28** after irradiation at 365 nm for 3 h.

• 8.4. Traceless

o Vinylsulphonylmethyl



- 1. RNH2
- 2. Chemistry
- 3. Alkylation
- 4. Elimination

recycled

8.5. Specific reagents/catalysts

SilyloxyBuLi and TFA stablenot TBAF (THF)

- 8.5. Special reagents/catalysts
 - o Oxidatively cleavable, Cu(II)acetate (diazene formation)
 TFA/piperidine stable
 - o Hydride cleavage

How to select the most appropriate...

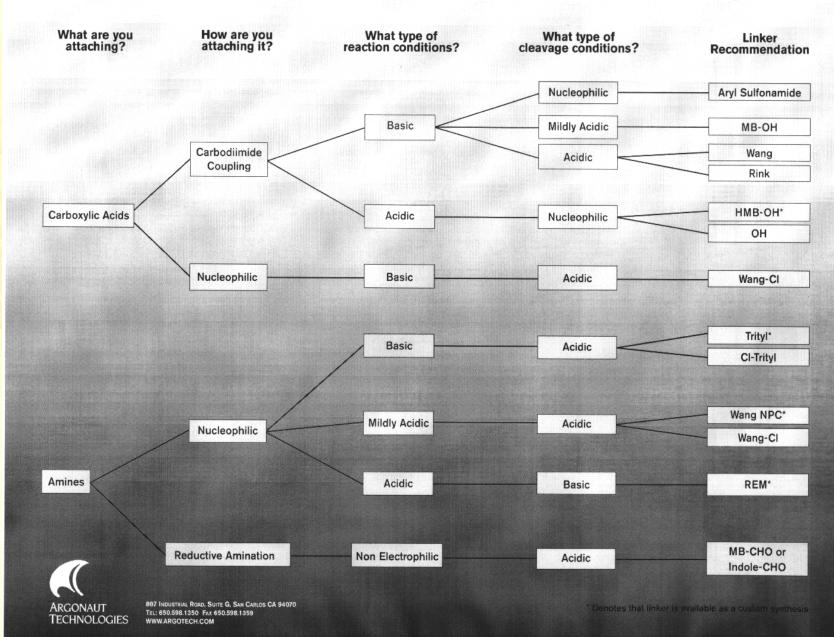
Support

- o Chemical compatibility
- Swelling compatibility
- Loading availability

How to select the most appropriate...

Solid Phase Linker Selection Guide

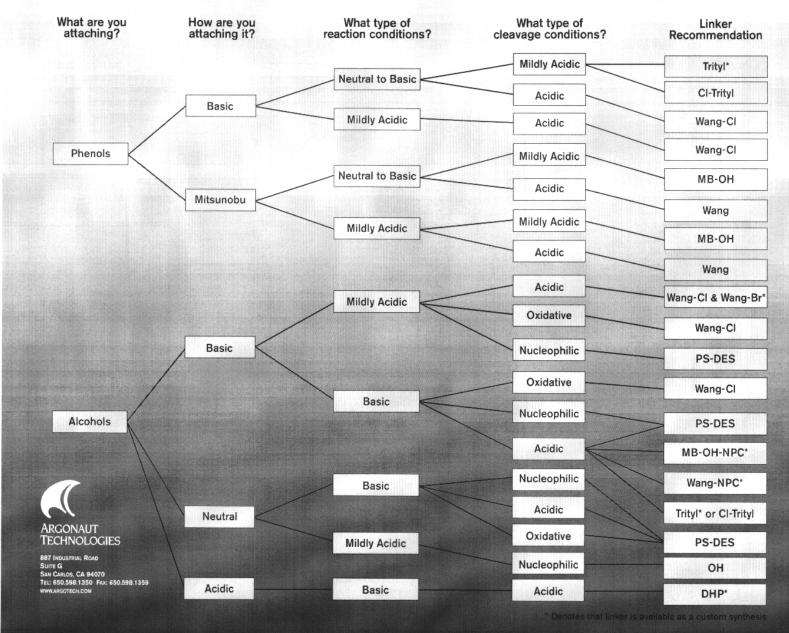
Linker



How to select the most appropriate...

Solid Phase Linker Selection Guide

Linker



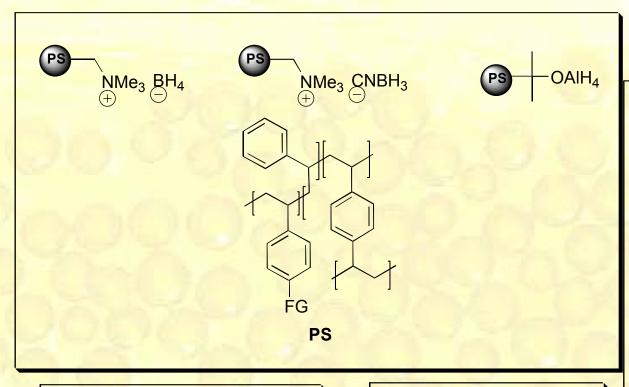
Lecture 4

Polymer-assisted solution phase synthesis

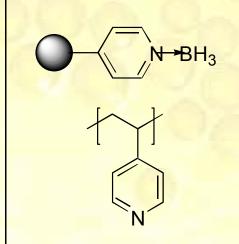
Combines the advantages of solution-phase synthesis (analysis, readily available products) with those of solid-phase synthesis (separation by filtration, use of excess of reagents).

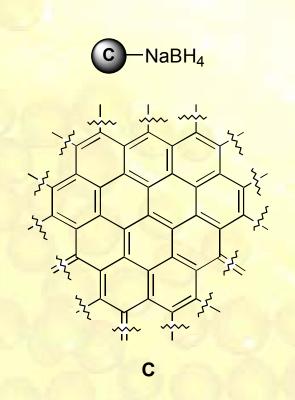
No need of linkers and cleavage protocols.

Polymer supported reducing agents



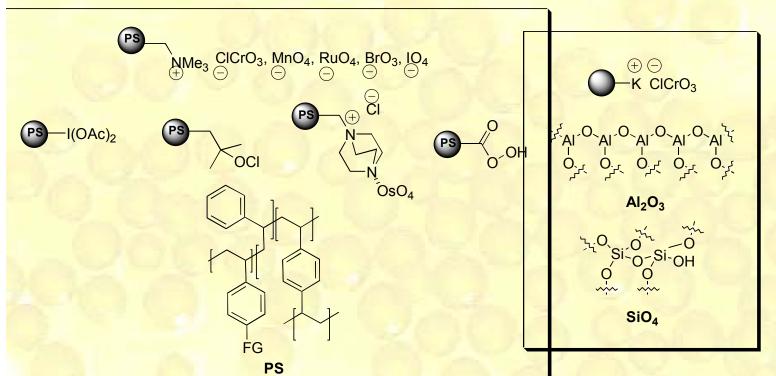
$$A = Al_2O_3$$





Polymer supported reducing agents

Polymer supported oxidising agents



Polymer supported oxidising agents

Polymers as reagents Polymer supported reagents for C-C bond formation